Antifungal Diterpenoid Alkaloids from Delphinium denudatum

Atta-ur-Rahman,* Amber Nasreen, Farzana Akhtar, M. Saleh Shekhani, Jon Clardy,[†] Masood Parvez,[‡] and M. Iqbal Choudhary

H. E. J. Research Institute of Chemistry, University of Karachi, Karachi-75270, Pakistan, Department of Chemistry, Cornell University, Ithaca, New York 14853-1301, and Department of Chemistry, The University of Calgary, Calgary, Alberta, Canada, T2N 1N4

Received September 25, 1996[®]

The roots of *Delphinium denudatum* have yielded a new diterpenoid alkaloid, 8-acetylheterophyllisine (1), in addition to the known alkaloids vilmorrianone (2), panicutine (3), denudatine (4), isotalatizidine (5), and condelphine (6), as well as 3-hydroxy-2-methyl-4*H*-pyran-4-one (7). Compounds 1, 3, and 7 have not been isolated from this plant previously. The structures of compounds 1, 3, and 7 were determined through spectral and X-ray diffraction analyses. Compounds 1-3 have shown antifungal activity against a number of human pathogenic fungi.

Delphinium species (Ranunculaceae) contain diterpenoid alkaloids that are generally of the veatchine or atisine type.¹ Delphinium species have also been used for the treatment of itches and other skin eruptions in folklore. Delphinium denudatum is widely distributed in the Western Himalayas and in Kashmir at altitudes of 2400-3650 m, especially on grassy slopes. The roots of this plant are bitter and used as a stimulant, alterative, and tonic.² Organic solvent extracts of the plant have been shown to have antimicrobial and immunomodulating properties.³ The radioprotectant effect of the aqueous extract of *D. denudatum* against radiation-induced changes in rat myocardium has also been investigated.⁴ The EtOH extracts of the roots of D. denudatum collected from Kashmir (Pakistan) have shown antifungal activity against Stachybotrys atra, Trichophyton longifusus, Curvularia lunata, Drechslera rostrata, Epidermophyton flocosum, Microsporum canis, Nigrospora oryzae, and Ganoderma applanatum,⁵ and antibacterial activity against Corynebacterium diphtheriae, Proteus vulgaris, Salmonella typhi, and Klebsiella pneumoniae.6

Continuing our work on the important medicinal plants of Pakistan, we wish to report here the isolation of a new diterpenoid alkaloid, 8-acetylheterophyllisine (1), from the roots of *D. denudatum*. Other compounds isolated from this plant were vilmorrianone (2), panicutine (3), denudatine (4), isotalatizidine (5), condelphine (6), and 3-hydroxy-2-methyl-4*H*-pyran-4-one (7). Compounds 1 and 2 showed antifungal activity against *Allescheria boydii, E. floccosum,* and *Aspergillus niger.*⁵ Panicutine (3) exhibited antifungal activity against *A. boydii, S. atra, Pleurotus ostreatus, N. oryzae, Dutarium rotatum,* and *A. niger.*⁵

The antifungal activity of compounds 1-3 was determined by the Agar tube diffusion method.⁶ Test tubes charged with sterile Sabouraud dextrose agar were inoculated with test compounds at different concentrations and were kept in a slanting position at room temperature. On solidification the test fungal cultures were inoculated on the slant, and growth inhibitions were observed after an incubation period of 7 days. Nystatin and griseofulvin were used as standard anti-



fungal drugs. MIC values of the antifungal compounds were determined by an agar tube dilution method⁵ and are presented in Table 1.

8-Acetylheterophyllisine (1), a heteratisine-type diterpenoid base, was isolated by column and thin-layer chromatographies. The HREIMS showed a molecular ion at m/z 417.2506 corresponding to the molecular formula $C_{24}H_{35}NO_5$, which required eight degrees of unsaturation. Compound 1 exhibited only terminal UV absorption. The IR spectrum of the compound exhibited a strong absorption band at 1720 (ester C=O) cm⁻¹. The ¹H-NMR spectrum of 1 indicated the presence of *N*CH₂-CH₃ (δ 0.99 and 2.89), OCH₃ (δ 3.24), and acetyl methyl (δ 1.95) groups.

Structure **1** was confirmed by an X-ray diffraction method. A suitable crystal that formed in the orthorhombic space group $P_{2_12_12_1}$ was selected for the study. Accurate lattice constants were a = 8.049(6) Å, b = 16.091(8) Å, and c = 16.964(7) Å. One molecule of composition $C_{23}H_{27}NO_5$ formed the asymmetric unit. All unique diffraction maxima with $2\theta \le 45^\circ$ were collected using 2θ : θ scans and graphite monochromated Mo K_a radiation (0.71069 Å). A total of 1686 unique reflections

[†] Cornell University.

[‡] The University of Calgary. [®] Abstract published in *Advance ACS Abstracts,* March 15, 1997.

Table 1. Evaluation of Antifungal Activity of Compounds **1**, **2**, and **3** against Pathogenic Fungi (MIC values in μ g/mL)^{*a*}

compds tested	A. boydii	A. niger	E. floccosum	P. ostreatus
1	100	200	250	150
2	150	100	225	175
3	75	125	200	125

 a Nystatin and griseofulvin were used as standards. Tests were carried out employing the agar tube dilution method. 5



Figure 1. Computer-generated perspective drawing of the final X-ray model of 8-acetylheterophyllisine (1).

were collected, 1400 (83%) were judged observed $[(|F_0|) \ge 6\sigma(|F_0|)]$ and used in subsequent calculations. The structure was phased using direct methods and refined using full-matrix least-square techniques with anisotropic heavy atoms and isotropic riding hydrogens to a conventional crystallographic residual of 0.041 ($R_W = 0.053$) for the observed data.^{7,8} A computer-generated drawing of the final X-ray model of 8-acetylheterophyllisine is given in Figure 1. Hydrogens are omitted for clarity, and no absolute configuration is implied.

Panicutine (3) (heterophylloidine), a hetidine-type base, was previously isolated from Aconitum paniculatum Lam. and A. heterophylloides.⁹⁻¹¹ We have now isolated this compound from D. denudatum as colorless crystals, and its molecular formula was derived as $C_{23}H_{29}NO_4$ from the HREIMS (M⁺, *m*/*z* 383.21977). The structure was unimbiguously established by the single crystal X-ray diffraction technique. Panicutine (3) was recrystallized from MeOH-iso-octane, and a suitable crystal was selected for further study. The crystal formed in the orthorhombic space group $P2_12_12_1$, was a = 8.492(7) Å, b = 13.60(2) Å, c = 18.21(2) Å, with one molecule $(C_{23}H_{29}NO_4)$ in the asymmetric unit. All unique diffraction maxima with $2\theta \leq 105^{\circ}$ were collected using 2θ : θ scans and graphite monochromated with Cu Kα radiation (1.54178 Å). A total of 1600 unique reflections were collected. Of these 1400 (87.5%) were judged obtained observed $[(|F_0|) \ge 3\sigma(F_0)]$ and used in further calculations. The structure was solved by the direct method (SHELX) and refined by full-matrix leastsquares techniques to a final discrepancy index of 0.073 $(\dot{R}_W = 0.096)$ for observed data.^{7,8} A computer-generated perspective drawing of the final X-ray model is given in Figure 2.

3-Hydroxy-2-methyl-4*H*-pyran-4-one (**7**) was also isolated from this plant. This compound was earlier isolated from larch trees (*Larix decidua*), pine needles, chicory, wood tars, and a variety of other natural sources.^{12,13} Compound **7** was obtained as large colorless crystals. Cell constants and an orientation matrix



Figure 2. Computer-generated perspective drawing of the final X-ray model of panicutine (**3**).



Figure 3. Computer-generated perspective drawing of the final X-ray model of 3-hydroxy-2-methyl-4*H*-pyran-4-one (7).

for data collection were obtained from a least-square refinement using the setting angles of 25 carefully centered reflections. This corresponded to a monoclinic $P2_1/a$ crystal with cell constants a = 7.075(2) Å, b = 36.010(11) Å, c = 7.168(2) Å, $\beta = 109.47(2)^{\circ}$ and three independent molecules in the asymmetric unit. A total of 3112 unique reflections were collected using Mo K α radiations, of which 1205 (38.7%) were judged observed $[(|F_0|) \ge 2\sigma(|F_0|)]$ and used in further calculations. The structure was solved by direct methods (SIR92) and refined by the full-matrix least-square technique to a final discrepancy index of 0.076 ($R_W = 0.077$) for observed data.^{7,8} Hydrogen atoms are included but not refined. A computer-generated perspective drawing of the final X-ray model is given in Figure 3.

Vilmorrianone (2), an atisine-type diterpenoid alkaloid, was previously isolated from *A. vilmorrianum.*¹³ Denudatine (4) was earlier isolated from *A. jinyan*gense¹⁴ and *D. denudatum*,¹⁵ while isotalatizidine (5) was also isolated from various *Aconitum* species^{16–19} and *D. denudatum.*²⁰ Condelphine (6), an aconitine base, was reported from *A. delphinifolium*, *A. japonicum*,^{21,22} and *D. denudatum.*²⁰ These four known compounds were isolated in the present investigation from the roots of *D. denudatum*.

Experimental Section

General Experimental Procedures. The purity of the samples was checked on precoated Si gel 254 plates. Solvents were freshly distilled before use. Melting points were determined on a Büchi 535 melting point apparatus. Optical rotations were determined on a Polaritronic polarimeter. The UV spectra were recorded in CH₃OH on a Shimadzu UV 240 instrument. The IR spectra were recorded on JASCO IRA-1 IR spectrophotometer. The ¹H-NMR spectra were recorded in CDCl₃ on a Bruker AMX-400 NMR spectrometer at 400 MHz, while the ¹³C-NMR spectra were recorded on the same instrument at 100 MHz, with TMS as internal standard. MS were measured on a JEOL HX-110 mass spectrometer.

X-ray diffraction studies (compounds 1, 3) were conducted on a R3M/V four-circled diffractometer (Siemens) connected with a MicroVax II computer (Digital) using Cu K α or Mo K α radiations. The X-ray structure of compound 7 was determined on a Rigaku AFCGS diffractometer with graphite monochromated Mo Ka radiation.

Plant Material. The roots of D. denudatum Wall. (2 kg) were collected from Azad Kashmir, Pakistan, in the months of July and August 1992.

Extraction and Isolation. A concentrated EtOH extract of the plant was dissolved in MeOH and filtered over a sintered funnel. The filtrate was again evaporated to a gum (4 g) and subjected to column chromatography using Si gel as stationary phase. The column was eluted with CHCl3-MeOH mixtures of increasing polarity. The fraction obtained on elution with CHCl₃-MeOH (95:5) contained compounds 1-7, which were separated through TLC.

8-Acetylheterophyllisine (1): recrystallized from *i*-C₈H₁₈-CH₂Cl₂ as brown crystals (3.4 mg); mp 152 °C; $[\alpha]_D$ 38.24° (MeOH), 0.0523 g; UV (MeOH) 314 (log ϵ 1.93), 338 (log ϵ 1.98) nm; IR (CHCl₃) 1714, 1727 cm⁻¹; ¹H NMR (CDCl₃) δ 1.03 (3H, br s, *N*-CH₂-CH₃), 1.95 (3H, s, COCH₃), 0.75 (3H, s, CH₃), 3.24 (3H, s, OCH₃), 4.69 (1H, br m, $W_{1/2} = 14.0$ H₂, H-13), 3.25 (1H, d, J =8 Hz, H-9), 3.35 (1H, δ , H-17); ¹³C NMR (CDCl₃) δ 84.6 (C-1), 26.5 (C-2), 37.2 (C-3), 34.8 (C-4), 49.5 (C-5), 27.1 (C-6), 48.6 (C-7), 87.1 (C-8), 41.4 (C-9), 42.1 (C-10), 29.7 (C-12), 75.2 (C-13), 172.0 (C-14), 28.8 (C-15), 29.5 (C-16), 60.7 (C-17), 26.4 (C-18), 56.5 (C-19), 48.9 (N-CH₂-CH₃), 13.4 (*N*-CH₂-*C*H₃), 22.3 (OCO*C*H₃), 55.6 (OCH₃), 169.8 (COCH₃); HREIMS m/z 417.2356 (M⁺, C₂₄H₃₅-NO₅, 6), 358.2374 (C₂₂H₃₂NO₃, 65), 326.2113 (C₂₁H₂₈-NO₂, 15).

Crystal Data for 1: $C_{23}H_{27}NO_5$, MW = 417.453, orthorhombic, $P2_12_12_1$, a = 8.049(6) Å, b = 16.091(8) Å, c = 16.964(7) Å, V = 2197(2) Å³, Z = 4, Dx = 1.262 mg/ cm³, Mo K α ($\lambda = 0.710$ 69 Å), F(000) = 904, T = 293 °K, R = 0.041, and Rw = 0.053, for 1400 unique observed reflections with $I > 2.00\sigma(I)$ (total = 1686), collected on a Rigaku AFC6S four-circuled diffracto meter using a crystal of approximate dimensions of $0.15 \times 0.15 \times 0.25$ mm to a maximum 2θ value of 50.0°. The structure was solved by direct methods are refined by full-matrix leastsquares calculations. The nonhydrogen atoms were refined anistropically.

Crystal Data for 3: $C_{18}H_{29}NO_4$, MW = 383.219, orthorhombic, $P2_12_12_1$ was a = 8.492(7) Å, b = 13.60(2)Å, c = 18.21(2) Å, V = 2102(4) Å³, Z = 4, Dx = 1.211mg/cm³, Cu K α (λ = 1.541 78 Å), F(000) = 824, T = 296 °K, R = 0.073, and RW = 0.096, for a total of 1600 unique reflections we judged od observed $[|F_0| \ge 3\sigma(F_0)]$ and used in further calculations. All the data were collected on a computer-controlled R3M/V four-circle diffractometer, using a crystal of approximate dimensions of 0.25 imes 0.22 imes 0.25 mm to a maximum 2heta value of 50.0°. The structure was solved by direct methods (SHELX) and refined by full-matrix least squares calculations. The nonhydrogen atoms were refined anistropically.

Crystal Data for 7: $C_6H_6O_3$, MW = 126.11, monoclinic P21/a was a = 7.075(2) Å, b = 36.010(11) Å, c =7.168(2) Å, $\beta = 109.47(2)^\circ$, V = 1721.8(a) Å³, Z = 12, D_X $= 1.459 \text{ mg/cm}^3$, Mo K α ($\lambda = 0.0710$, 69 Å), F (000) = 792.00, T = -136.0 °K, R = 0.076, and $R_W = 0.077$, for 1205 unique observed reflections with $I > 2.00\sigma(I)$ (total 3112), collected on a Rigaku AFC6S diffractometer using a crystal of approximate dimensions of 0.40 \times 0.40 \times 0.33 mm. The structure was solved by direct methods (SIR 92) refined by full-matrix least-squares techniques.

Additional crystallographic details (atomic coordinates and equivalent isotropic displacement coefficients, lists of structure factors, interatomic distances, angles, anisotropic displacement coefficients for non-hydrogen atoms, hydrogen atom coordinates and their isotropic displacement co-efficients) of 1, 3, and 7 have been deposited at the Cambridge Crystallographic Data Centre. The coordinates can be obtained, upon request, from Dr. Olga Kennord, University Chemical Laboratory, 12 Union Road, Cambridge CB2 1EZ, UK.

Acknowledgment. The work at Cornell University was partially supported by NIH Grant No. CA24487 to J. C. Two authors (A.R. and M.I.C.) are grateful to the Highnoon Laboratories (Pvt.), Lahore, Pakistan, for travel grants. We are also grateful to the INFAQ Foundation for financial support to F. A.

References and Notes

- (1) Atta-ur-Rahman. Handbook of Natural Products Data, Diterpenoid and Steroidal Alkaloids; Elsevier Science Publishers: Amsterdam, 1990.
- (2) Bhatnagar, S. S. The Wealth of India; CSIR: New Delhi, Vol. III, 1952; p 30.
- (3) Siddiqui, R.; Kazmi, S. U.; Shekhani, S. Proceedings of the 90th Annual Meeting of the American Society for Microbiology, Anaheim, California, May 13–17, 1990.
- (4) Khan, A. B. J. Mol. Cell. Cardiol. 1989, 21, S171.
- (5) Paxton, J. D. Methods in Plant Biochemistry; Waterman, P. G., Ed. Academic Press: London, 1991, Vol. 6; pp 33-51. Carran, R.; Maran, A.; Montero, J. M.; Fernandolago, L.;
- (6)Dominaguez, A. Plantes Med. Phytother. 1987, 21, 195-202.
- (7) Archival crystallographic data have been deposited with the Cambridge Crystallographic Data Center, University Chemical Laboratory, Lensfield Road, Cambridge CB2 1EW, UK
- (8) Sheldrick, R. G. SHELXTL PLUS, Structure Solving Package, Nicolet (now Siemens), Madison, WI, 1986
- (9) Pelletier, S. W.; Joshi, B. S.; Desai, H. K.; Al Panu, A.; Katz, A. Heterocycles 1986, 24, 1275-1277.
- (10) Katz, A.; Staehelin, E. *Helv. Chim. Acta* 1982, *65*, 286–289.
 (11) Pelletier, S. W.; Mody, M. V.; F. Moore, J.; Desai, H. K.; Puri, H. S. *Tetrahedron Lett.* 1981, *22*, 313–314.
- (12) J. Buckingham, Ed. Dictionary of Natural Products; Chapman & Hall: London, Vol. 3, 1994; p 3112.
 (13) Ding, L.; Chen, Y.; Wu, F. *Planta Med.* 1991, *57*, 275–277.
- (14) Chen, D.; Sung, W. L. Yaoxue Xuebao 1981, 16, 748-751.
- (15) Götz, M.; Wiesner, K. Tetrahedron Lett. 1969, 50, 4369-4372.
- (16) Bando, H.; Wada, K.; Amiya, T.; Kobayashi, K.; Fujimoto, Y.; Sakurai, T. *Heterocycles* **1987**, *26*, 2623–2637.
- (17) Gonzalez, A. G.; Fuente, G.; Orribo, T.; Acosta, R. D. Heterocycles 1985, 23, 2979-2982.
- (18) Boido, V.; Edwards, O. E.; Handa, K. L.; Kolt, R. J.; Purushothaman, K. K. Can. J. Chem. 1984, 62, 778-784.
- (19) Konno, C.; Shirasaka, M.; Hikino, H. J. Nat. Prod. (Lloydia), 1982, 45, 128-133.
- (20) Pelletier, S. W.; Keith, L. H.; Parthasarathy, P. C. J. Am. Chem. Soc. 1967, 89, 4146-4157
- (21) Aiyar, V. N.; Kulanthaivel, P.; Benn, M. Phytochemistry 1986, *25*, 973–975.
- (22) Sakai, S.; Takayama, H.; Okamoto, T. Yakugaku Zasshi 1979, 99. 647-656.

NP960663N